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Synthetic studies on bafilomycin A_1 : stereoselective synthesis of the $C_{12}-C_{17}$ fragment and its coupling with the C_1-C_{11} subunit

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Abstract—Stereoselective addition of (*E*)-1-lithio-2-tributylstannylethylene on a chiral cyclic di-*t*-butylsilyleneketal C_{14} – C_{17} aldehyde afforded the required Felkin–Anh adduct for the synthesis of the C_{12} – C_{17} fragment of bafilomycin A₁, the configuration of which was assigned unambiguously. After appropriate coupling with the enantiopure C_1 – C_{11} fragment, the C_{12} – C_{17} subunit obtained here can be used for the study of the 16-membered macrolide formation either by an acyl activation or an intramolecular Stille reaction. Intermolecular esterification of the 15-OH with an acyl activation of the carboxylic acid of the C_1 – C_{11} fragment, in modified Yamaguchi's conditions, affords here an intermediate for examining an intramolecular Stille coupling. © 2004 Elsevier Ltd. All rights reserved.

In the preceding communication,¹ we described the synthesis of the enantiopure C_1 - C_{11} fragment **1** of bafilomycin A_1 . We herein report the synthesis of the C_{12} - C_{17} subunit **2** and its structure assignment, and also the preparation of **3** (Scheme 1), in order to examine the formation of the 16-membered macrolactone of bafilomycin A_1 via an intramolecular Stille coupling, which will be discussed in the accompanying communication.^{2,3}

1. Synthesis of the C₁₂-C₁₇ fragment

In a first approach, we prepared the enantiopure aldehyde 4,³ expecting that the OTBS silyl ether, α to the aldehyde, might disfavour the formation of the Cramchelate adduct with lithio derivatives (or other organometallics) and allow to get the desired stereoisomer stereoselectively.⁴ However, the addition of the E-lithiovinyltributylstannane 6, generated in situ from 5^{5} was found to be almost nonstereoselective in THF/hexane (Scheme 2). The configurations of the two adducts, 7 and 8, were demonstrated unambiguously by a chemical filiation as discussed further. Modifications of reaction conditions (temperature, excess of 6, addition of BF₃·Et₂O in THF or LiClO₄ in Et₂O) were unsatisfactory. Results were not improved with the parent OTES aldehyde 9. Additions of TMS-C=C-Li (1.5 equiv, THF, 0°C, overnight) and n-BuLi (1.2 equiv, toluene, -78 °C, 6 h) were also weakly stereoselective with 4, affording 55/45 and 65/35 mixtures of two diastereoisomers, respectively, moreover in analogous moderate yields.³ The low stereoselectivity obtained in all those condensations is most likely due to the fact that the two



Scheme 1.

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Scheme 2. Reagents and conditions: (a) 6 (1.05 equiv), addition of 4 in THF, -78 °C, 4 h, then MeOH quench at -78 °C; (b) 6 (1.05 equiv), addition of 10 in THF, -78 °C, 7 h, then MeOH quench at -78 °C.

substituents, α to the aldehyde, are large and probably result in a dynamic equilibrium of conformers involved in the reaction; the steric hindrance resulting from these two substituents is also probably responsible of the moderate reactivity of the aldehyde **4**.

We then examined the condensations of **6** with the cyclic di-*t*-butylsilyleneketal aldehyde **10** in order to still have a nonchelating group and also to see if the cyclic ketal might favour a conformation of the aldehvde. In fact the best results were thus obtained with 10 (in a 95/5 ratio with the epimerized aldehyde, see further) in THF since the adducts **11** and **12** were isolated in 40% and 10.5% yield, respectively, in quasi stoichiometric conditions (Scheme 2); a mixture of two other diastereoisomers was also isolated in 6% yield and some pure starting aldehyde 10 was still recovered (13.5%) after chromatography. These results were obtained reproducibly at a 0.2-3.5 mmole scale. The configurations of 11 and 12 were demonstrated by a chemical filiation, as shown further. Yields of adducts 11 and 12 were lower and stereoselection was not significantly improved either with DME (-78 °C, 6h) or THF/HMPA (9/1) (-78 °C, 4h); a 50/50 mixture of the two same adducts was obtained with LiClO₄ (2 equiv) in Et₂O (-78 °C, 3.5 h).

On the other hand, the aldehyde 13 gave with 6, at -78 °C, a 50/50 mixture of the corresponding adducts in THF or with LiClO₄ (2 equiv) in Pr₂O.³



The aldehyde 10 was prepared from commercially available cis-4-benzyloxy-2-buten-1-ol 14 (Scheme 3). Sharpless asymmetric epoxidation⁶ afforded the epoxyalcohol 15 in 76% yield with an ee of 80%, determined by ¹H NMR (400 MHz) of the corresponding MTPA esters.7 We then preferred to open directly the epoxide by Me₂CuLi in Et₂O, even if the reaction afforded a 70/ 30 mixture of the two regioisomers as in previous related examples,⁸ rather than to develop a more lengthy sequence to solve that problem. As the two isomers 16 and 17 were not separable, the most efficient solution was to use the crude mixture for converting them into the corresponding di-t-butylsilyleneketals since those corresponding to 1,2-diols are known to be very labile.⁹ Thus after aqueous work-up and chromatography over silicagel, the desired ketal 18 was isolated in 58% yield over the two steps from 15. After hydrogenolysis of the benzyl ether (84% yield), Swern oxidation¹⁰ of the alcohol 19, using *i*-Pr₂NEt to minimize the epimerization of this sensitive aldehyde, afforded 10 in 90% yield as a 95/5 mixture of epimers at C_{15} , which could not be



Scheme 3. Reagents and conditions: (a) $Ti(O-i-Pr)_4$ (1.0 equiv), (-)-DET (1.2 equiv), anhyd CH_2Cl_2 , -30 °C, 30 min, then 14, -30 °C, 1 h, followed by *t*-BuOOH (2.6 M in anhyd CH_2Cl_2 , 5 equiv, -30 °C, 4 days); (b) CuI (5 equiv), MeLi 1.6 M in Et₂O (9 equiv) at 0 °C, then -40 °C, 15, 5 h; (c) crude mixture of 16 and 17 (70/30), 2,6-lutidine (10 equiv), *t*-Bu₂Si(OTf)₂ (3.2 equiv), CH₂Cl₂, rt, 3 h; (d) 10% Pd/C (40% in weight), H₂, 95% EtOH, rt, 4 h; (e) oxalyl chloride (1.2 equiv), DMSO (2.4 equiv), CH₂Cl₂, -78 °C, 15 min, then 19, 30 min, followed by *i*-Pr₂NEt (5.0 equiv), -78 °C to rt, 1 h.

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separated. It is worthy to point out that epimerization was observed only on upscaling, after chromatography (heptane/AcOEt 95/5). The configuration of the aldehyde **10** was clearly demonstrated by NMR (${}^{3}J_{15,16} = 4.5$ Hz, compared to ${}^{3}J_{15,16} = 11$ Hz for the minor diastereoisomer). The aldehyde **10** is thus obtained in 33% overall yield from **14** (five steps).

The configurations of the adducts 7, 8, 11 and 12, were determined unambiguously by the chemical filiation between them and 22, which was an intermediate of the synthesis of bafilomycin A_1 achieved by Toshima et al. (Scheme 4).¹¹ The NMR spectra of the acetonides 22 and 25 are clearly different in CDCl₃ (¹H at 300 MHz, ¹³C at 50 MHz), and comparison with the data published by Toshima for 22 allows an unambiguous assignment of the structures.¹²

Formation of the methyl ether at C_{14} was quite a difficult problem for avoiding silyl migration, desilylation or destannylation for the adducts 7 and 11, and showed to be even more difficult for 8 and 12. The corresponding methyl ethers could however be obtained by reaction of the derived lithium alkoxide, generated in situ, with a large excess of methyl iodide, in THF/HMPA at room temperature. Fortunately, yields were higher with the diastereoisomer required further for the synthesis. Evans and Calter already pointed out the difficulty of the formation of the methyl ether for a related compound with the (*S*) configuration at C_{14} , and gave an other solution to this problem.¹³ The di-*t*-butylsilylene ketal could be cleaved with an excess TBAF in THF, at room temperature, with no destannylation, 20 thus affording the diol 21 in 88% yield. The diols 21 and 24 were then converted into the corresponding acetonides 22 and 25 with 2-methoxypropene and PPTS as a catalyst, conditions which were mild enough to avoid protodestannylation. The chemical filiations of 11 with 7, and of 12 with 8, were achieved via 2 and 26, respectively.³

For the synthesis of bafilomycin A_1 , the required C_{12} - C_{17} fragment **2** is thus obtained with a 10% overall yield from **14** (ee = 80%, nine steps).

2. Precursor of the macrolide formation (Scheme 5)

The prepared subunits 1 and 2 now allowed us to study the formation of the macrolide either via an acyl activation of their coupling product obtained after an intermolecular Stille coupling, or via an intramolecular Stille reaction of the corresponding ester 3. For bafilomycin A1 synthesis, the 16-membered macrolide formation has always been achieved via an acyl activation.11,13-15 Therefore, instead of an acyl activation and in order to find an alternate solution, we decided to examine the formation of the macrocycle via an intramolecular Stille coupling.¹⁶ Thus, the required enantiopure precursor 3 was obtained in 89% yield by an intermolecular esterification of the enantiopure C1-C11 acid 1 with the chiral C_{12} - C_{17} alcohol 2 (ee = 80%) (1.08 equiv), in the presence of Yamaguchi's chloride,¹⁷ NEt₃ and DMAP, at room temperature, and 11% of the



Scheme 4. Reagents and conditions: (a) *n*-BuLi 1.6 M in hexane (1.2 equiv), MeI (20 equiv), THF, -78 °C to rt, 1 h, then HMPA to get THF/HMPA (10/1), rt, 16 h; (b) TBAF 1 M in THF (5 equiv), rt, 3 days; (c) CH₂=CMe(OMe) (3 equiv), PPTS (0.01 equiv), CH₂Cl₂, rt, 20 h; (d) DMTBF₄ (1.8 equiv for **21**, 1.1 equiv for **24**), 2,6-di-*t*-butyl-4-methylpyridine (2.5 equiv for **21**, 1.5 equiv for **24**), MeCN, rt, 4 h; (e) *n*-BuLi 1.6 M in hexane (1.2 equiv), MeI (100 equiv), THF, -78 °C to rt, 1 h, then HMPA to get THF/HMPA (5/1), rt, 16 h; (f) TBAF 1 M in THF (10 equiv), rt, 5 days for **2**, 6 days for **26**.



Scheme 5. Reagents and conditions: 1, toluene, DMAP (2 equiv), rt, then addition of NEt₃ (4 equiv), followed by 2,4,6-trichlorobenzoyl chloride (2 equiv) and addition of 2 in toluene (1.08 equiv), then rt, 24 h.

alcohol **2** (ee not determined) were reisolated after chromatography (Scheme 5).¹⁸

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- Compound 11: pale yellow oil; [α]_D -190 (*c* 0.05, CHCl₃);
 IR (cm⁻¹, CHCl₃): 3598 (OH), 1598 (C=C); ¹H NMR (300 MHz, CDCl₃): δ/TMS 6.29 (m, 2H, H₁₂, H₁₃), 4.05

(m, 1H, H₁₄), 4.10 (dd, 1H, H₁₅, $J_{15,16} = 2.5$, $J_{15,14} = 8$), 4.34 (dd, 1H, H_{17a} , $J_{17a,17b} = 11$, $J_{17a,16} = 3$), 3.92 (dd, 1H, H_{17b} , $J_{17a,17b} = 11, J_{17b,16} = 2), 1.81$ (d, 1H, OH, $J_{OH,14} = 4), 2.01$ (m, 1H, H₁₆), 1.48 (m, 6H, CH₃CH₂CH₂CH₂Sn), 1.30 (m, 6H, $CH_3CH_2CH_2CH_2Sn$), 1.22 (d, 3H, CH_3 , $J_{16,CH_3} = 7$), 1.07 (s, 9H, t-BuSi), 1.05 (s, 9H, t-BuSi), 0.90 (t, 6H, $CH_3CH_2CH_2CH_2Sn$, $J_{CH_2CH_2Sn} = 7$), 0.88(t, 9H. $CH_3CH_2CH_2CH_2Sn, J_{CH_3CH_2} = 7$); ¹³C NMR (50.3 MHz, CDCl₃): 149.0 (C₁₃), 127.5 (C₁₂), 78.8 (C₁₅), 74.9 (C₁₄), 71.4 (C17), 34.2 (C16), 29.2 and 27.4 (CH3CH2CH2CH2Sn), 28.7 and 27.6 (t-BuSi), 23.5 and 20.7 (CqSi), 13.8 $(CH_3CH_2CH_2CH_2Sn)$, 11.4 (CH_3) , 9.5 $(CH_3CH_2CH_2)$ CH_2 Sn); $C_{27}H_{56}O_3$ SiSn = 575.53; MS (EI, m/z): 575 (M⁺), 518, 461.

Compound 12: pale yellow oil; $[\alpha]_D + 2.0$ (c 0.4, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ /TMS 6.41 (d, 1H, H₁₂, $J_{13,12} = 19$), 5.88 (dd, 1H, H₁₃, $J_{13,12} = 19$, $J_{13,14} = 7$), 4.00 (m, 1H, H₁₄), 4.05 (dd, 1H, H₁₅, $J_{15,16} = 2.5$, $J_{15,14} = 8$), 4.33 (dd, 1H, H_{17a} , $J_{17a,17b} = 11$, $J_{17a,16} = 3$), 3.89 (dd, 1H, H_{17b} , $J_{17a,17b} = 11, J_{17b,16} = 2$, 2.98 (d, 1H, OH, $J_{OH,14} = 1$), 1.78 (m, 1H, H₁₆), 1.47 (m, 6H, CH₃CH₂CH₂CH₂Sn), 1.29 (m, 6H, $CH_3CH_2CH_2CH_2Sn$), 1.16 (d, 3H, CH_3 , $J_{16,CH_3} = 7$), 1.10 (s, 9H, t-BuSi), 1.06 (s, 9H, t-BuSi), 0.89 (t, 6H, $CH_3CH_2CH_2CH_2Sn, \quad J_{CH_2CH_2Sn} = 7),$ 0.87(t, 9H. $CH_3CH_2CH_2CH_2Sn$, $J_{CH_3CH_2} = 7$); ¹³C NMR (50.3 MHz, CDCl₃): 144.9 (C₁₃), 133.4 (C₁₂), 79.3 (C₁₅), 77.2 (C₁₄), 71.0 (C₁₇), 34.1 (C₁₆), 29.0 and 27.1 (CH₃CH₂CH₂CH₂Sn), 28.5 and 27.4 (t-BuSi), 23.3 and 20.6 (CqSi), 13.6 (CH₃CH₂CH₂CH₂CH₂Sn), 11.5 (CH₃), 9.3 (CH₃CH₂-CH₂CH₂Sn); C₂₇H₅₆O₃SiSn = 575.53; MS (EI, m/z): 575 (M^+) , 518.

Compound 22: pale yellow oil; IR (cm⁻¹, CHCl₃): 1600 (C=C); ¹H NMR (300 MHz, CDCl₃): δ /TMS 6.21 (d, 1H, H_{12} , $J_{12,13} = 19$), 5.79 (dd, 1H, H_{13} , $J_{12,13} = 19$, $J_{13,14} = 6$), 4.08 (dd, 1H, H_{17a} , $J_{17a,17b} = 11.5$, $J_{17a,16} = 3$), 3.82 (dd, 1H, $H_{15}, J_{15,14} = 9, J_{15,16} = 2$, 3.60 (dd, 1H, $H_{17b}, J_{17a,17b} = 11.5$, $J_{17b,16} = 2$), 3.40 (dd, 1H, H₁₄, $J_{14,15} = 9$, $J_{14,13} = 6$), 3.28 (s, 3H, OCH₃), 1.77 (m, 1H, H₁₆), 1.37 (s, 3H, CH_{3gem}), 1.35 (s, 3H, CH_{3gem}), 1.48 (m, 6H, CH₃CH₂CH₂CH₂Sn), 1.30 (m, 6H, $CH_3CH_2CH_2CH_2Sn$), 1.10 (d, 3H, CH_3 , $J_{16,CH_3} =$ 7), 0.89 (t, 6H, CH₃CH₂CH₂CH₂CH₂Sn, $J_{CH_2CH_2Sn} = 7$), 0.88 (t, 9H, CH₃CH₂CH₂CH₂CR, $J_{CH_2CH_2} = 7$); ¹³C NMR 9H, $CH_3CH_2CH_2CH_2Sn$, $J_{CH_3CH_2} = 7$); (50.3 MHz, CDCl₃): 146.2 (C₁₃), 132.7 (C₁₂), 98.7 (Cq), 83.6 (C₁₄), 73.8 (C₁₅), 67.1 (C₁₇), 56.4 (OCH₃), 30.4 and 29.7 (CH3gem), 29.6 and 27.3 (CH3CH2CH2CH2Sn), 19.0 (C₁₆), 13.8 (CH₃CH₂CH₂CH₂Sn), 11.2 (CH₃), 9.6 $(CH_3CH_2CH_2CH_2Sn); C_{23}H_{46}O_3Sn = 489.32; MS$ (EI, m/z): 490 (M⁺).

Compound **25**: pale yellow oil; IR (cm⁻¹, CHCl₃): 1601 (C=C); ¹H NMR (300 MHz, CDCl₃): δ /TMS 6.33 (d, 1H, H₁₂, J_{12,13} = 19), 5.66 (dd, 1H, H₁₃, J_{12,13} = 19, J_{13,14} = 8.5), 4.11 (dd, 1H, H_{17a}, J_{17a,17b} = 11.5, J_{17a,16} = 2.5), 3.91 (dd, 1H, H₁₅, J_{15,14} = 8.5, J_{15,16} = 2.5), 3.55 (dd, 1H, H_{17b}, J_{17a,17b} = 11.5, J_{17b,16} = 2), 3.49 (dd, 1H, H₁₄, J_{14,15} = 1, J_{14,13} = 8.5), 3.29 (s, 3H, OCH₃), 1.48 (s, 3H, CH_{3gem}), 1.47 (m, 6H, CH₃CH₂CH₂CH₂Sh), 1.46 (s, 3H, CH_{3gem}), 1.43

(m, 1H, H₁₆), 1.32 (m, 6H, CH₃*CH*₂CH₂CH₂Sn), 1.07 (d, 3H, CH₃, $J_{16,CH_3} = 7$), 0.92 (t, 6H, CH₃CH₂CH₂*CH*₂Sn, $J_{CH_2CH_2Sn} = 7$), 0.88 (t, 9H, *CH*₃CH₂CH₂CH₂Sn, $J_{CH_3CH_2} = 7$); ¹³C NMR (50.3 MHz, CDCl₃): 143.1 (C₁₃), 136.0 (C₁₂), 99.0 (Cq), 86.8 (C₁₄), 74.0 (C₁₅), 67.2 (C₁₇), 56.4 (OCH₃), 30.4 and 29.9 (CH_{3gem}), 29.2 and 27.2 (CH₃*CH*₂*CH*₂CH₂Sn), 19.0 (C₁₆), 13.8 (*CH*₃CH₂-CH₂CH₂Sn), 10.9 (CH₃), 9.6 (CH₃CH₂-CH₂CH₂Sn), 10.9 (CH₃), 9.6 (CH₃CH₂CH₂CH₂Sn); C₂₃H₄₆O₃Sn = 489.32; MS (EI, *m/z*): 490 (M⁺).

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- 18. Compound **3**: pale yellow oil; $[\alpha]_D + 27$ (*c* 0.7, CHCl₃); IR (cm⁻¹, CHCl₃): 1710 (C=O), 1607 (C=C), 1509 (Ar); ¹H NMR (300 MHz, CDCl₃): δ /TMS 7.43 (d, 2H, Ha, $J_{\text{Ha,Hb}} = 9$), 7.31 (d, 4H, Hd, $J_{\text{Hd,He}} = 9$), 7.26 (m, 2H, Hb), 7.17 (m, 1H, Hc), 6.81 (d, 4H, He, $J_{\text{Hd,He}} = 9$), 6.51 (s, 1H, H₃), 5.98 (d, 1H, H₁₂, $J_{13,12} = 19$), 5.89 (dq, 1H, H₅,

 $J_{5,6} = 10, J_{CH_{3},5} = 1), 5.83$ (s, 1H, H₁₁), 5.83 (dd, 1H, H₁₃), $J_{13,12} = 19$, $J_{13,14} = 6$), 5.38 (dd, 1H, H₁₅, $J_{15,14} = 6$, $J_{15,16} = 6$), 3.77 (s, 6H, OCH₃), 3.50 (m, 1H, H₁₄), 3.49 (s, 3H, CH₃, C₂OCH₃), 3.39 (dd, 1H, H₇, $J_{6,7} = 3.5$, $J_{7.8} = 5.5$), 3.21 (s, 3H, CH₃, HC₁₄OCH₃), 3.03 (dd, 1H, H_{17a} , $J_{17a,17b} = 9$, $J_{17a,16} = 6$), 2.92 (dd, 1H, H_{17b} , $J_{17a,17b} = 9$, $J_{17b,16} = 6$), 2.68 (m, 1H, H₆), 2.44 (dd, 1H, H_{9a}, $J_{9a,9b} = 13$, $J_{9a,8} = 3$), 2.20 (m, 1H, H₁₆), 1.94 (d, 3H, CH₃, $J_{CH_{3},5} = 1$), 1.92 (m, 1H, H_{9b}), 1.78 (s, 3H, CH₃), 1.72 (m, 1H, H₈), 1.45 (m, 6H, CH₃CH₂CH₂CH₂Sn), 1.27 (m, 6H, $CH_3CH_2CH_2CH_2Sn$), 1.04 (d, 3H, CH_3 , $J_{16,CH_3} = 7$), 0.96 (d, 3H, CH₃, $J_{6,CH_3} = 7$), 0.96 (t, 9H, CH_3CH_2Si , $J_{CH_3,CH_2Si} = 8$), 0.86 (t, 9H, $CH_3CH_2CH_2CH_2Sn$, $J_{CH_3CH_2} = 7$; t, 6H, CH₃CH₂CH₂CH₂Sn, $J_{CH_2CH_2Sn} = 7$), 0.73 (\tilde{d} , 3H, CH₃, $J_{8,CH_3} = 7$), 0.66 (q, 6H, CH₃CH₂Si, $J_{CH_3,CH_2Si} = 8$; ¹³C NMR (50.3 MHz, CDCl₃): 164.2 (C₁), 158.1 (C=COCH₃), 146.9 (C₁₀), 144.7 (C₁₃), 144.4 and 136.2 (Cq Ar), 142.9 and 135.2 (C2, C4), 140.9 (C5), 130.2 (C₃), 129.3(C₁₂), 129.9, 128.1, 127.5 and 126.4 (CH Ar), 112.8 (C=COCH₃), 85.4 (C₁₅), 85.2 (CqO), 80.4 (C₇), 75.4 (C_{11}) , 75.0 (C_{14}) , 65.7 (C_{17}) , 60.0 (C_2OCH_3) , 56.1 (HC₁₄OCH₃), 54.9 (OCH₃), 42.9 (C₉), 35.8 and 34.7 (C₆, C₈), 28.9 and 27.0 (CH₃CH₂CH₂CH₂Sn), 24.5, 18.2, 15.6, 14.5 and 13.5 (CH₃), 13.9 (CH₃CH₂CH₂CH₂Sn), 12.6 $(CH_3CH_2CH_2CH_2Sn)$, 6.9 (CH_3) , 5.3 (CH_2Si) ; $C_{63}H_{97}IO_8SiSn = 1256.15$; MS (FD, m/z): 1256 (M⁺), 1199, 1128.